

APPENDICES

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่

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APPENDIX A

CHEMICALS AND REAGENTS

Chemical or Reagent	Source
1 Kb Plus DNA Ladder™	Gibco BRL, NY, USA
Acetic acid, glacial	Merck, Darmstadt, Germany
Agar granulated	BBL, MI, USA
Ammonium acetate	Fluka, Buchs, Switzerland
Ampicillin	Sigma, MO, USA
Boric acid	Merck, Darmstadt, Germany
Bromophenol blue	Merck, Darmstadt, Germany
D (+) Glucose	Fluka, Buchs, Switzerland
Disodium Ethylenediaminetetra acetate·2H ₂ O	Amresco, OH, USA
Ethidium bromide	Sigma, MO, USA
Ethyl alcohol	Merck, Darmstadt, Germany
FACS Lysing Solution	Becton Dickinson, CA, USA
Glycerol	Sigma, MO, USA
High Pure PCR Template Preparation Kit	Roche Molecular Biochemicals, Mannheim, Germany
High Pure Plasmid Isolation Kit	Roche Molecular Biochemicals Mannheim, Germany
<i>Hinc</i> II restriction endonuclease	Promega, USA
Hydrochloric acid	Merck, Darmstadt, Germany
Isoprep	Robbins Scientific co., CA, USA
Isopropanol	Merck, Darmstadt, Germany

LE agarose	Seakem-BMA, ME, USA
Magnesium chloride hexahydrate	Merck, Darmstadt, Germany
Magnesium sulfate heptahydrate	Merck, Darmstadt, Germany
MinElute Gel Extraction Kit	Qiagen, Germany
pGEM®-T Easy Vector Kit	Promega, USA
Platinum® <i>Taq</i> DNA Polymerase High Fidelity	Invitrogen, USA
Potassium acetate	Merck, Darmstadt, Germany
Potassium chloride	Merck, Darmstadt, Germany
Potassium dihydrogen phosphate	Merck, Darmstadt, Germany
Sodium chloride	Merck, Darmstadt, Germany
Sodium dodecyl sulfate	Sigma, MO, USA
Sodium hydrogen phosphate	Merck, Darmstadt, Germany
Sodium hydroxide	Merck, Darmstadt, Germany
SureFill™ 6% Sequencing gel	Visible Genetic, USA
<i>Taq</i> DNA polymerase	Promega, USA
Thermo Sequenase Cy5 Dye Terminator Cycle Sequencing Kit	Amersham Bioscience, England
Tris base	Sigma, MO, USA
Tryptone	Difco Laboratories, MI, USA
Yeast extract	Gibco, NY, USA

APPENDIX B

INSTRUMENTS

Instrument-model	Source
Analytical balance	Sartorius, Germany
Biological safety cabinet class II	Gelman Sciences, Australia
Dry Block Heater	Biosan Laboratory Inc., USA
Flow cytometer-FACSCalibur	Beckton Dickinson, USA
Gel Toaster™ unit	Visible Genetic, USA
GeneObjects™ software	Visible Genetic, USA
Hemocytometer counting chamber	Improved Neubauer, USA
Horizontal Gel Electrophoresis System	BRL, USA
Hot Air Oven	Sheldon, USA
Incubator shaker	Amerex Instruments Inc., USA
Light microscope	Olympus, Japan
Long-Read Tower™ sequencer	Visible Genetic, USA
MicroCel 700 Cassette	Visible Genetic, USA
Refrigerated centrifuge	Sorvall, Germany
Refrigerated microcentrifuge	Sorvall, Germany
Refrigerator (4°C)	Sanyo, Thailand
Refrigerator (-20°C)	Sanyo, Thailand
Refrigerator (-70°C)	Kelvinator Scientific, USA
pH Meter	Pierce, USA
Photo Documentation	Fotodyne, USA

Primer Premier version 5.00 software	PREMIER Biosoft International, CA, USA
Sero-fuge centrifuge	Beckton Dickinson, USA
Spectrophotometer UV1201	Shimadzu Co., Japan
SureFill™ Injector	Visible Genetic, USA
Thermal Cycler GeneAmp® PCR system 2700	Applied Biosystems, USA
Ultraviolet transilluminator	Vilber Lourmat, France
Vortex Mixer	Scientific Industries, USA
Water bath	Sheldon, USA

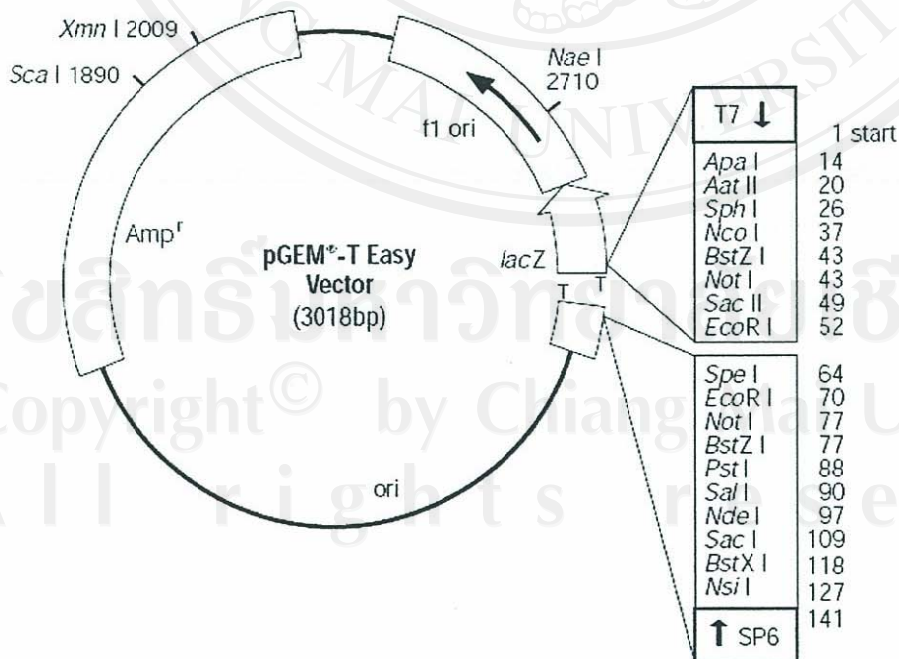


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APPENDIX C

pGEM®-T Easy Vector

The pGEM®-T Easy Vector System is a convenient system for the cloning of PCR products. The vector is prepared by cutting pGEM®-T Easy Vectors with *EcoR* V at base 60 of the sequence and adding a 3' terminal thymidine to both ends. These single 3'-T overhangs at the insertion site greatly improve the efficiency of ligation of a PCR product into the plasmids by preventing recircularization of the vector and providing a compatible overhang for PCR products generated by certain thermostable polymerases. These polymerases often add a single deoxyadenosine, in a template-independent fashion, to the 3'-ends of the amplified fragments.

1. pGEM®-T Easy Vector sequence circle map

2. pGEM®-T Easy Vector Sequence reference points

T7 RNA Polymerase transcription initiation site	1
SP6 RNA Polymerase transcription initiation site	141
T7 RNA Polymerase promoter	3002-6
SP6 RNA Polymerase promoter	136-158
multiple cloning site	10-128
lacZ start codon	180
lacoperon sequences	2839-2999, 166-395
lacoperator	100-216
b-lactamase coding region	1337-2197
phage fl region	2383-2838
binding site of pUC/M13 Forward Sequencing Primer	2959-2975
binding site of pUC/M13 Reverse Sequencing Primer	176-192

3. pGEM®-T Easy Vector Sequence

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1 GGGCGAATTG GGCCCGACGT CGCATGCTCC CGGCCGCCAT GGCGGCCGCG
51 GGAATTCGAT*ATCACTAGTG AATTCGCGGC CGCCTGCAGG TCGACCATAT
101 GGGAGAGCTC CCAACGCGTT GGATGCATAG CTTGAGTATT CTATAGTGTC
151 ACCTAAATAG CTTGGCGTAA TCATGGTCAT AGCTGTTTCC TGTGTGAAAT
201 TGTTATCCGC TCACAATTCC ACACAACATA CGAGCCGGAA GCATAAAGTG
251 TAAAGCCTGG GGTGCCTAAT GAGTGAGCTA ACTCACATTA ATTGCGTTGC
301 GCTCACTGCC CGCTTTCCAG TCGGGAAACC TGTCGTGCCA GCTGCATTAA
351 TGAATCGGCC AACGCGCGGG GAGAGGCGGT TTGCGTATTG GGCGCTCTTC
401 CGCTTCCTCG CTCACTGACT CGCTGCGCTC GGTCGTTCGG CTGCGGCGAG
451 CGGTATCAGC TCACTCAAAG GCGGTAATAC GGTTATCCAC AGAATCAGGG
501 GATAACGCAG GAAAGAACAT GTGAGCAAAA GGCCAGCAA AGGCCAGGAA
551 CCGTAAAAAG GCCGCGTTGC TGGCGTTTTT CCATAGGCTC CGCCCCCTG
601 ACGAGCATCA CAAAAATCGA CGCTCAAGTC AGAGGTGGCG AAACCCGACA
651 GGACTATAAA GATACCAGGC GTTTCCCCCT GGAAGCTCCC TCGTGCCTC
701 TCCTGTTCCG ACCCTGCCGC TTACCGGATA CCTGTCCGCC TTTCTCCCTT
751 CGGGAAGCGT GGCCTTTTCT CATAGCTCAC GCTGTAGGTA TCTCAGTTCCG
801 GTGTAGGTCG TTCGCTCCAA GCTGGGCTGT GTGCACGAAC CCCCCGTTCA
851 GCCCGACCGA TGCGCCTTAT CCGGTAACTA TCGTCTTGAG TCCAACCCGG
901 TAAGACACGA CTTATCGCCA CTGGCAGCAG CCACTGGTAA CAGGATTAGC
951 AGAGCGAGGT ATGTAGGCGG TGCTACAGAG TTCTTGAAGT GGTGGCCTAA
1001 CTACGGCTAC ACTAGAAGGA CAGTATTGG TATCTGCGCT CTGCTGAAGC
1051 CAGTTACCTT CGGAAAAAGA GTTGGTAGCT CTTGATCCGG CAAACAAACC

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1101 ACCGCTGGTA GCGGTGGTTT TTTTGGTTGC AAGCAGCAGA TTACGCGCAG
 1151 AAAAAAAGGA TCTCAAGAAG ATCCTTTGAT CTTTTCTACG GGGTCTGACG
 1201 CTCAGTGGAA CGAAAACTCA CGTTAAGGGA TTTTGGTCAT GAGATTATCA
 1251 AAAAGGATCT TCACCTAGAT CCTTTTAAAT TAAAAATGAA GTTTTAAATC
 1301 AATCTAAAGT ATATATGAGT AAACCTGGTC TGACAGTTAC CAATGCTTAA
 1351 TCAGTGAGGC ACCTATCTCA GCGAICTGTC TATTTTCGTTT ATCCATAGTT
 1401 GCCTGACTCC CCGTCGTGTA GATAACTACG ATACGGGAGG GCTTACCATC
 1451 TGGCCCCAGT GCTGCAATGA TACCGCGAGA CCCACGCTCA CCGGCTCCAG
 1501 ATTTATCAGC AATAAACCCAG CCAGCCGGAA GGGCCGAGCG CAGAAGTGGT
 1551 CCTGCAACTT TATCCGCCCTC CATCCAGTCT ATTAATTGTT GCCGGGAAGC
 1601 TAGAGTAAGT AGTTCGCCAG TTAATAGTTT GCGCAACGTT GTTGGCATTG
 1651 CTACAGGCAT CGTGGTGTCA CGCTCGTCGT TTGGTATGGC TTCATTACAGC
 1701 TCCGGTCCC AACGATCAAG GCGAGTTACA TGATCCCCCA TGTGTGCAA
 1751 AAAAGCGGTT AGCTCCTTCG GTCCITCCGAT CGTTGTCAGA AGTAAGTTGG
 1801 CCGCAGTGTT ATCACTCATG GTTATGGCAG CACTGCATAA TTCTCTTACT
 1851 GTCATGCCAT CCGTAAGATG CTTTTCTGTG ACTGGTGAGT ACTCAACCAA
 1901 GTCATTCTGA GAATAGTGTA TGC GCGGACC GAGTTGCTCT TGCCCGGCGT
 1951 CAATACGGGA TAATACCGCG CCACATAGCA GAACTTTAAA AGTGCTCATC
 2001 ATTGGAAAAC GTTCTTCGGG GCGAAAACTC TCAAGGATCT TACCGCTGTT
 2051 GAGATCCAGT TCGATGTAAC CCACTCGTGC ACCCAACTGA TCTTCAGCAT
 2101 CTTTTACTTT CACCAGCGTT TCTGGGTGAG CAAAAACAGG AAGGCAAAT
 2151 GCCGCAAAAA AGGGAATAAG GCGGACACGG AAATGTTGAA TACTCATACT
 2201 CTTCTTTTT CAATATTATT GAAGCATTTA TCAGGGTTAT TGTCTCATGA
 2251 GCGGATACAT ATTTGAATGT ATTTAGAAAA ATAAACAAAT AGGGGTTCCG
 2301 CGCACATTTT CCCGAAAAGT GCCACCTGTA TGCGGTGTGA AATACCGCAC
 2351 AGATGCGTAA GGAGAAAATA CCGCATCAGG CGAAATTGTA AACGTTAATA
 2401 TTTTGTAAA ATTCGCGTTA AATATTTGTT AAATCAGCTC ATTTTTTAAAC
 2451 CAATAGGCCG AAATCGGCAA AATCCCTTAT AAATCAAAG AATAGACCGA
 2501 GATAGGGTTG AGTGTGTGTC CAGTTTGGAA CAAGAGTCCA CTATTAAAGA
 2551 ACGTGGACTC CAACGTCAA GGGCGAAAAA CCGTCTATCA GGGCGATGGC
 2601 CCACTACGTG AACCATCACC CAAATCAAGT TTTTTCGCGT CGAGGTGCCG
 2651 TAAAGCTCTA AATCGGAACC CTAAAGGGAG CCCCCGATTT AGAGCTTGAC
 2701 GGGGAAAGCC GCGAACGTG GCGAGAAAGG AAGGGAAGAA AGCGAAAGGA
 2751 GCGGGCGCTA GGGCGCTGGC AAGTGTAGCG GTCACGCTGC GCGTAACCAC
 2801 CACACCCGCC GCGCTTAATG CGCCGCTACA GGGCGCGTCC ATTCGCCATT
 2851 CAGGCTGCGC AACTGTTGGG AAGGGCGATC GGTGCGGGCC TCTTCGCTAT
 2901 TACGCCAGCT GCGGAAAGGG GGATGTGCTG CAAGGCGATT AAGTTGGGTA
 2951 ACGCCAGGGT TTTCCAGTC ACGACGTTGT AAAACGACGG CCAGTGAATT
 3001 GTAATACGAC TCACTATA

* Cloned insert site indicated by an asterisk

APPENDIX D

NUCLEIC ACID AND AMINO ACID SYMBOL

1. Nucleic acid symbol

Nucleic acid	Symbol
Adenine	A
Cytocine	C
Guanine	G
Inosine	I
Thymine	T
Uracil	U
A + C	M
A + G	R
A + T	W
C + G	S
C + T	Y
G + T	K
A + C + G	V
A + C + T	H
A + G + T	D
C + G + T	B
A + C + G + T	N

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2. Amino acid symbol

Amino acid	Three letter abbreviation	One letter symbol
Alanine	Ala	A
Arginine	Arg	R
Asparagine	Asn	N
Aspartic acid	Asp	D
Asparagine or aspartic acid	Asx	B
Cysteine	Cys	C
Glutamine	Gln	Q
Glutamic acid	Glu	E
Glutamine or glutamic acid	Glx	Z
Glycine	Gly	G
Histidine	His	H
Isoleucine	Ile	I
Leucine	Leu	L
Lysine	Lys	K
Methionine	Met	M
Phenylalanine	Phe	F
Proline	Pro	P
Serine	Ser	S
Threonine	Thr	T
Tryptophan	Trp	W
Tyrosine	Tyr	Y
Valine	Val	V

APPENDIX E

REAGENTS AND BUFFERS PREPARATION

1. Commonly used stock solutions

1 M Tris-HCl (pH 8.0)

Tris base	121.1 g
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- Dissolve in 800 ml distilled water
- Adjust the pH to 8.0 by adding concentrated HCl
- Adjust the volume to 1 liter with distilled water
- Sterilize by autoclaving and store at room temperature

0.5 M EDTA (pH 8.0)

Disodium Ethylenediaminetetra acetate·2H ₂ O	186.1 g
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- Dissolve in 800 ml distilled water
- Adjust the pH to 8.0 by adding NaOH
- Adjust the volume to 1 liter with distilled water
- Sterilize by autoclaving and store at room temperature

5 M Potassium acetate (pH 7.5)

Potassium acetate	49.1 g
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- Dissolve in 90 ml distilled water
- Adjust the pH to 7.5 by adding acetic acid
- Adjust the volume to 100 ml with distilled water
- Sterilize by autoclaving and store at 4°C

10 % SDS

Sodium dodecyl sulfate	1 ml
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Distilled water	100 ml
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2. Reagents for isolation of the PBMCs

Phosphate buffer saline (PBS) pH 7.2

NaCl	8 g
KCl	0.2 g
Na ₂ HPO ₄	1.44 g
KH ₂ PO ₄	0.24 g

- Dissolve in 800 ml distilled water
- Adjust the pH to 7.2 with HCl and fill up distilled water to 1 liter
- Sterilize by autoclaving and store at room temperature

TE buffer (pH 8.0)

Tris-HCl (pH 8.0)	10 mM
EDTA (pH 8.0)	1 mM

- Sterilize by autoclaving and store at room temperature

3. Reagents for Electrophoresis

50X Tris-acetate Buffer (TAE)

Tris base	242 g
Glacial acetic acid	57.1 ml
0.5 M EDTA (pH 8.0)	100 ml

- Dissolve in 1 liter distilled water
- Sterilize by autoclaving and store at room temperature

5X Tris-borate Buffer (TBE)

Tris base	54 g
Boric acid	27.5 g
0.5 M EDTA (pH 8.0)	20 ml

- Dissolve in 1 liter distilled water
- Sterilize by autoclaving and store at room temperature

6X gel loading buffer

Bromophenol blue	0.25 %
Glycerol	30 %

- Mixed thoroughly and keep at -20°C

10 mg/ml Ethidium bromide

Ethidium bromide 1 g

- Dissolve in 100 ml distilled water
- Stir on a magnetic stirrer to ensure that the dye has dissolved
- Wrap the container in aluminum foil and store at room temperature

1% agarose gel

LE agarose 1 g

0.5X TAE 100 ml

2% agarose gel

LE agarose 2 g

0.5X TAE 100 ml

3% agarose gel

LE agarose 3 g

0.5X TAE 100 ml

1 Kb Plus DNA Ladder

1 µg/µl 1 Kb Plus DNA Ladder™ (Gibco BRL, USA) 5 µl

6X gel loading buffer 10 µl

Distilled water 45 µl

4. Reagents for the cloning of PCR products**2 M Mg²⁺ stock**

MgCl₂ · 6H₂O 20.33 g

MgSO₄ · 7H₂O 24.65 g

- Dissolved in 100 ml distilled water
- Sterilize by filtration and store at room temperature

2 M glucose

Glucose 36 g

Distilled water 100 ml

- Sterilize by filtration

SOC medium

Tryptone	2.0 g
Yeast extract	0.5 g
1 M NaCl	1 ml
1 M KCl	0.25 ml
2 M Mg ²⁺ stock	1 ml
2 M glucose	1 ml

- Add tryptone, yeast extract, NaCl and KCl to 97 ml distilled water and stir to dissolve
- Autoclave and cool to room temperature
- Add 2M Mg²⁺ stock and 2M glucose, each to a final concentration of 20 mM
- Fill up sterile distilled water to 100 ml
- Sterilize by passage the complete medium through a 0.2 µm filter unit and store at 4°C

LB medium

Tryptone	10 g
Yeast extract	5 g
NaCl	5 g

- Dissolved in 800 ml distilled water
- Adjust pH to 7.0 with NaOH and fill up distilled water to 1 liter
- Sterilize by autoclaving and store at 4°C

LB plates with ampicillin

Agar granulated	15 g
LB medium	1 liter

- Sterilize by autoclaving and allow the medium to cool to 50°C
- Add ampicillin to a final concentration of 100 µg/ml
- Pour 30-35 ml of medium into 85 mm Petri dishes and let the agar harden
- Store at 4°C for up to 1 month

5. Reagent for minipreparations of plasmid DNA**Solution I**

Glucose	50 mM
Tris-HCl (pH 8.0)	25 mM
EDTA (pH 8.0)	10 mM

- Sterilize by autoclaving and store at 4°C

Solution II

NaOH (freshly diluted from 10N stock)	0.2 N
SDS	1 %

Solution III

5M Potassium acetate	60 ml
Glacial acetic acid	11.5 ml
Distilled water	28.5 ml

7.5 M Ammonium acetate

Ammonium acetate	77.75 g
Distilled water	100 ml

- Sterilize by filtration and store at 4°C

70 % ethanol

Absolute ethanol	70 ml
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- Add distilled water up to 100 ml

CURRICULUM VITAE

Name Tanawan Samleerat

Date of birth August 13, 1979

Place of birth Bangkok, Thailand

Education

1996 Certificate of senior high school
Suksanari high school, Bangkok

2000 Bachelor of Science in Medical
Technology, B.Sc. (Med. Tech.)
Faculty of Allied Health Sciences
Thammasat University

Scholarships Received

2002 Graduate Research Scholarships,
Ministry of University Affairs, Thailand

Publications

Samleerat, T. Role of Cytokines/Chemokines in AIDS: Disease Progression and Resistance (Review). Bull Chiang Mai Assoc Med Sci 2001; 34(2): 107-117.

Samleerat, T., Sritana, N., Dettrairat, S., Kunachiwa, W. and Leechanachai, P. Study of Mutation in CCR5 Gene; $\Delta 32$, -m303 and Its Protein Density Expressed on the Surface of CD4+ Lymphocytes from HIV-1-Highly Exposed Persistently Seronegative Persons.

International Symposium on New Developments in Biological Monitoring and Clinical Management of HIV Infection. The Empress Hotel, Chiang Mai, Thailand. May 24-26, 2002 (poster presentation).

Samleerat, T., Sritana, N., Dettrairat, S., Kunachiwa, W. and Leechanachai, P. Study of Mutation in CCR5 Gene; $\Delta 32$, -m303 and Its Protein Density Expressed on the Surface of CD4+ Lymphocytes from HIV-1-Highly Exposed Persistently Seronegative Persons. The 3rd National Symposium on Graduate Research, Suranaree University of Technology, Nakhon Ratchasima, Thailand. July 18, 2002 (oral presentation).

Leechanachai, P., **Samleerat, T.,** Dettrairat, S. and Kunachiwa, W. Polymorphisms of CCR5 gene and its' regulatory region in HIV-1 Highly Exposed Persistently Seronegative (HEPS) group. The 2nd IAS Conference on HIV Pathogenesis and Treatment at the Palais des Congres. Paris, France. July 13-16, 2003 (poster presentation).