

APPENDIX A

CHEMICALS AND REAGENTS

| Name of chemical/ reagent | Source/ Company |
|--------------------------------------|--|
| Absolute ethanol | Merck, Darmstadt, Germany |
| Acetic acid | Merck, Darmstadt, Germany |
| Acrylamide | Bio-Rad, Richmond, CA, USA |
| Agarose | Sigma-Aldrich, St. Louis, MO, USA |
| Ammonium persulfate (APS) | Bio-Rad, Richmond, CA, USA |
| Bis (N, N'-Methylene-bis-acrylamide) | Bio-Rad, Richmond, CA, USA |
| Bovine serum albumin | PIERCE, Rockford, IL, USA |
| Bromphenol blue | Sigma-Aldrich, St. Louis, MO, USA |
| Commercial grade curcuminoids | Sigma-Aldrich, St. Louis, MO, USA |
| Coomasie brilliant blue R-250 | Sigma-Aldrich, St. Louis, MO, USA |
| Copper sulfate | Merck, Darmstadt, Germany |
| Developer and replenisher | Kodak, NY, USA |
| Diethyl pyrocarbonate (DEPC) | Sigma-Aldrich, St. Louis, MO, USA |
| Dimethyl sulfoxide (DMSO) | Sigma-Aldrich, St. Louis, MO, USA LAB-SCAN, Bangkok, Thailand |
| Disodium hydrogen phosphate | Merck, Darmstadt, Germany Fluka, Buchs, Switzerland |
| DNA ladder | |
| - 1 kb DNA Ladder | GIBCO-BRL, Grand Island, NY, USA |
| - O'GeneRuler™ 1 kb DNA Ladder | MBI, Fermentas, Vilnius, Lithuania |
| - O'GeneRuler™ 100 bp DNA Ladder | MBI, Fermentas, Vilnius, Lithuania |
| Ethidium bromide | Sigma-Aldrich, St. Louis, MO, USA |
| EDTA (Disodium) | Merck, Darmstadt, Germany |

| | |
|--|------------------------------------|
| Fetal bovine serum | GIBCO-BRL, Grand Island, NY, USA |
| Folin & Cocalteu's phenol reagent | Merck, Darmstadt, Germany |
| Glycerol | Merck, Darmstadt, Germany |
| Glycine | Amresco, Ohio, USA |
| HEPES | Sigma-Aldrich, St. Louis, MO, USA |
| HRP conjugated goat anti-rabbit IgG | PIERCE, Rockford, IL, USA |
| Hydrochloric acid (HCl) | Merck, Darmstadt, Germany |
| Isopropanol | Merck, Darmstadt, Germany |
| L-glutamine | GIBCO-BRL, Grand Island, NY, USA |
| Mercaptoethanol | Sigma-Aldrich, St. Louis, MO, USA |
| Methanol | LAB-SCAN, Bangkok, Thailand |
| MTT | Sigma-Aldrich, St. Louis, MO, USA |
| NE-PER [®] Nuclear and Cytoplasmic Extraction reagents | PIERCE, Rockford, IL, USA |
| PageBlue [™] Protein Staining Solution | MBI, Fermentas, Vilnius, Lithuania |
| PageRuler [™] Prestained Protein Ladder | MBI, Fermentas, Vilnius, Lithuania |
| Penicillin-streptomycin | GIBCO-BRL, Grand Island, NY, USA |
| Potassium chloride | Merck, Darmstadt, Germany |
| Potassium dihydrogen phosphate | Merck, Darmstadt, Germany |
| Primers | Invitrogen, California, USA |
| Rabbit polyclonal anti-GAPDH antibody (GAPDH; FL-335) | PIERCE, Rockford, IL, USA |
| Rabbit polyclonal anti-GAPDH antibody (WT1; C-19) | PIERCE, Rockford, IL, USA |
| Restore [™] Western Blot Stripping Buffer | PIERCE, Rockford, IL, USA |
| RPMI-1640 powder | GIBCO-BRL, Grand Island, NY, USA |
| Skim milk | Fluka, Buchs, Switzerland |
| Sodium bicarbonate | Merck, Darmstadt, Germany |
| Sodium carbonate | Merck, Darmstadt, Germany |
| Sodium chloride | Merck, Darmstadt, Germany |

| | |
|---|-----------------------------------|
| Sodium dodecyl sulfate (SDS) | Sigma-Aldrich, St, Louis, MO, USA |
| Sodium potassium tartrate | Sigma-Aldrich, St, Louis, MO, USA |
| Sucrose | Sigma-Aldrich, St, Louis, MO, USA |
| SuperScript TM III One-Step RT-PCR System With Platinum [®] Tag DNA Polymerase | Invitrogen, California, USA |
| SuperSignal [®] West Pico Chemiluminescent Substrate | PIERCE, Rockford, IL, USA |
| TEMED | Bio-Rad, Richmond, CA, USA |
| Tris hydrochloride (Tris-HCl) | Sigma-Aldrich, St, Louis, MO, USA |
| TRIzol TM reagent | Invitrogen, California, USA |
| Trypan blue | Sigma-Aldrich, St, Louis, MO, USA |
| Tween 20 | Sigma-Aldrich, St, Louis, MO, USA |

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APPENDIX B**INSTRUMENTS**

| Instrument | Company |
|-----------------------------------|--|
| Analytical balance | OHAUS CORPORATION |
| Autoclave | HUXLER |
| Automatic pipette | BIOHIT, GILSON, LABMATE, and BIO-RAD |
| Automatic pipette tip | BIOLINE |
| Carbon dioxide incubator | JOUAN |
| Centrifuge | CLAY ADAMS |
| 15 or 50 ml centrifuge tube | GREINER BIO-ONE AND CORNING INCORPORATION |
| Deionized distilled water machine | PK WATER TEXT |
| ECL-hyper film | PIERCE |
| Freezer (-80°C) | PTW ULTRA COLD |
| Freezer (-20°C) | SANYO |
| Gel documentation | BIO-RAD |
| Gel electrophoresis apparatus | HANGZHOU BIOER TECHNOLOGY |
| 10 cm glass plate | PYREX and PETRIO |
| Homogenizer | PARGUS (JAPAN) |
| Hot air oven | THAI STAINLESS ARGON |
| Inverted microscope | OLYMPUS |
| Kodak medical X-ray Film | KODAK |
| Laminar flow biological cabinet | CLEAN |
| Light microscope | OLYMPUS |

| | |
|---|---------------------------------|
| Nitrocellulose membrane | PALL CORPORATION |
| Magnetic stirrer | SYBRON/ THERMOLYNE |
| Microcentrifuge, bench-topped | EPPENDORF |
| Microcentrifuge tube | CLP |
| Milipore filter paper | PALL CORPORATION |
| Mini protein II slab gel | BIO-RAD |
| Pipet-aid | DRUMMOND |
| Pasture pipette | PYREX |
| RCR amplifier | EPPENDORF |
| pH meter | THERMO ORION |
| Power supply | E-C APPARATUS CORPORATION |
| Refrigerator | TOSHIBA |
| Serological pipette | PYREX |
| Spectrophotometer | SHIMADZU, BARA SCIENTIFIC |
| 25 or 75 cm ³ T-flask | NUNC |
| Thin-wall PCR tube | MOLECULAR BIO PRODUCTS |
| Trans-blot [®] electrophoretic transfer cell | BIO-RAD |
| Vortex mixer | GEMMY INDUSTRIAL CORPORATION |
| Water bath | GFL |

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APPENDIX C

REAGENTS AND BUFFER PREPARATION

Human leukemic cell lines culture

1. Incomplete RPMI 1640 medium

| | | |
|---------------------------|------|---------------|
| RPMI powder (GIBCO BRL) | 10.4 | g (1 package) |
| HEPES | 3.57 | g |
| NaHCO ₃ | 2.0 | g |
| 0.34% 2-mercaptoethanol | 1.0 | mL |
| Deionized distilled water | 800 | mL |

Adjust pH to 7.2-7.4 then adjust volume to 1,000 mL and sterilized by suction filter (membrane pore size 0.2 μ m), stored at 4°C and checked sterility before used.

2. Complete RPMI 1640 medium

| | | |
|-----------------------------|------|----|
| Incomplete RPMI 1640 medium | 88.5 | mL |
| Fetal bovine serum | 10.0 | mL |
| L-glutamine | 1.0 | mL |
| Pen/strep | 0.5 | mL |

Stored at 4°C

3. Freezing solution

8% DMSO in fetal bovine serum

| | | |
|--------------------|-----|----|
| Fetal bovine serum | 9.2 | mL |
| DMSO | 0.8 | mL |

Stored at 4°C

4. Phosphate buffer saline (PBS) pH 7.4

| | | |
|----------------------------------|------|---|
| KH ₂ PO ₄ | 0.24 | g |
| Na ₂ HPO ₄ | 1.44 | g |
| NaCl | 8.0 | g |
| KCl | 0.2 | g |

Dissolve in 800 mL deionized distilled water, adjusted pH to 7.4 then top up to 1,000 mL and sterilized by autoclave.

Cell survival measurement

1. MTT stock dye solution

| | | |
|------------|-----|----|
| MTT | 1.0 | g |
| PBS pH 7.4 | 200 | mL |

After dissolve MTT dye, filtrate any nonsoluble powder by filtration with membrane filterpore size 0.2 μm , collected in dark container.

2. 0.2% (w/v) trypan blue

| | | |
|-------------|-----|----|
| Trypan blue | 0.2 | g |
| PBS | 100 | mL |

Protein determination

1. Reagent A

| | | |
|---|------|----|
| 2% (w/v) Na_2CO_3 in 0.1 N NaOH | | |
| NaOH | 2.0 | g |
| Na_2CO_3 | 10.0 | g |
| Deionized distilled water | 500 | mL |

2. Reagent B

0.5% (w/v) $\text{CuSO}_4 \cdot 5 \text{H}_2\text{O}$ & 1% (w/v) $\text{NaKC}_4\text{H}_4\text{O}_6 \cdot 2\text{H}_2\text{O}$ (Na-K Tartrate)

Two reagents, CuSO_4 and Na-K Tartrate, were prepared as follow:

Part A: 0.5% (w/v) $\text{CuSO}_4 \cdot 5 \text{H}_2\text{O}$

| | | |
|--|-----|----|
| $\text{CuSO}_4 \cdot 5 \text{H}_2\text{O}$ | 0.5 | g |
| Deionized distilled water | 50 | mL |

Before using 0.5 mL of part A and part B were mixed.

3. Reagent C

Working solution was freshly prepared by mixing reagent A 50 mL and reagent B ratio 50:1

4. Folin-ciocalteau phenol reagent 1N

Folin- ciocalteau phenol reagent 2N was diluted in deionized distilled water to 1N.

SDS-PAGE analysis**1. Stock solution A: separating gel buffer 1.5 mM Tris-HCl, pH 8.8**

| | | |
|---------------------------|-------|----|
| Tris-base | 18.15 | g |
| Deionized distilled water | 80 | mL |

Adjust pH to 8.8 then adjust volume to 100 mL and filtrate any nonsoluble powder by filtration with membrane filter pore size 0.2 μ m, collect in dark container.

2. Stock solution C: stock acrylamide solution (30% T, 2.7%)

| | | |
|---------------------------|------|----|
| Acrylamide | 29.2 | g |
| Bis (Estaman) | 0.8 | g |
| Deionized distilled water | 70 | mL |

Adjust volume to 100 mL and filtrate any nonsoluble powder by filtration with membrane filter pore size 0.2 μ m, collect in dark container.

3. Stock solution D: stacking gel buffer 0.5 mM Tris HCl, pH 6.8

| | | |
|---------------------------|------|----|
| Tris base | 6.05 | g |
| Deionized distilled water | 70 | mL |

Adjust pH to 6.8 then adjust volume to 100 mL and filtrate any nonsoluble powder by filtration with membrane filter pore size 0.2 μ m, collect in dark container.

4. Electrode buffer (Running buffer)

| | | |
|-----------|------|---|
| Tris-base | 3.0 | g |
| Glycine | 14.4 | g |
| SDS | 1.0 | g |

Dissolve in deionized distilled water 1,000 mL then filtrate by suction filter and store at 4°C.

5. 5X non-reducing buffer

| | | |
|------------------------|-------|----|
| 1.0 M Tris-HCl, pH 6.8 | 0.625 | mL |
| Glycerol | 1.0 | mL |
| 1% Bromphenol blue | 0.125 | mL |

Adjust volume to 10 mL with distilled water.

6. 5X reducing buffer

| | | |
|------------------------|-----|---------|
| 5X non-reducing buffer | 475 | μ L |
| 2-mercaptoethanol | 25 | μ L |

7. Coomassie blue stain

| | | |
|----------------|------|----|
| Coomassie blue | 0.25 | g |
| Methanol | 20 | mL |
| Acetic acid | 10 | mL |

Deionized water was top up to 100 mL.

8. Coomassie blue destaining solution

| | | |
|-------------|-----|----|
| Methanol | 100 | mL |
| Acetic acid | 50 | mL |

Deionized water was top up to 500 mL.

9. Stock ammonium persulfate solution

10% (w/v) APS in deionized distilled water

| | | |
|---------------------------|-----|----|
| Ammonium persulfate | 0.1 | g |
| Deionized distilled water | 1.0 | mL |

10. Stock 10% SDS solution

| | | |
|---------------------------|-----|----|
| SDS | 0.2 | mL |
| Deionized distilled water | 1.0 | mL |

11. Separating gel 12%

| | | |
|-------------------------------|-----|---------|
| Deionized distilled water | 3.5 | mL |
| Tris HCl, pH 8.8 (solution A) | 2.5 | mL |
| 10% SDS | 100 | μ L |
| Acrylamide/Bis (solution C) | 4.0 | mL |
| 10% APS | 50 | μ L |
| TEMED | 5 | μ L |

12. Stacking gel 4%

| | | |
|-------------------------------|------|---------|
| Deionized distilled water | 3.05 | mL |
| Tris-HCl, pH 6.8 (solution D) | 1.25 | mL |
| 10% SDS | 50 | μ L |
| Acrylamide/Bis (solution C) | 0.65 | mL |
| 10% APS | 25 | μ L |
| TEMED | 5 | μ L |

Western blot analysis**1. Transferring buffer (Blotting buffer)**

| | | |
|-----------|------|----|
| Tris-base | 3.03 | g |
| Glycine | 14.4 | g |
| Methanol | 200 | mL |

Dissolve in deionized distilled water 1,000 mL then filtrate by suction filter and store at 4°C

2. Phosphate buffer saline (PBS) pH 7.4

| | | |
|----------------------------------|-------|---|
| NaH ₂ PO ₄ | 0.204 | g |
| Na ₂ HPO ₄ | 1.3 | g |
| NaCl | 7.28 | g |

Dissolve in 800 mL deionized distilled water, adjusted pH to 7.4 then adjusted volume to 1,000 mL and sterilized by autoclave or filtration.

3. Blocking reagent

| | | |
|-----------|----|----|
| Skim milk | 5 | g |
| Anti-foam | 20 | μL |

Dissolve in 100 mL PBS, pH 7.4

4. Washing buffer

| | | |
|-------------|-----|----|
| PBS, pH 7.4 | 500 | mL |
| Tween 20 | 500 | μL |

Reverse transcriptase polymerase chain reaction (RT-PCR)**1. Stock 0.5 M EDTA, pH 8.0**

| | | |
|--------------------|-------|---|
| EDTA | 186.1 | g |
| DEPC treated water | 1 | L |

Sterilized by autoclave

2. DEPC treated water

| | | |
|-----------------------------|-----|----|
| Deionized distilled water | 4 | L |
| Diethylpyrocarbonate (DEPC) | 400 | μL |

Shake it vigorously and store at room temperature for overnight and removed DEPC by autoclave.

3. 1 mg/mL Ethidium bromide

| | |
|--------------------|---------|
| Ethidium bromide | 0.001 g |
| DEPC treated water | 1.0 mL |

Stored at 4°C in dark.

4. 6X loading dye

| | |
|--------------------|----------|
| Bromphenol blue | 0.0025 g |
| Sucrose | 0.4 g |
| DEPC treated water | 1.0 mL |

Sucrose should be dissolved before adding phenol blue and then stored at 4°C in dark.

5. 50X TAE (Tris acetate buffer)

| | |
|---------------------|---------|
| Tris-base | 24.2 g |
| Gracial acetic acid | 5.71 mL |
| 0.5 M EDTA, pH 8.0 | 10 mL |
| DEPC trated water | 100 mL |

Stored at room temperature

6. 1% Agarose

| | |
|------------------|--------|
| Agarose | 1.0 g |
| 1X TAE | 100 mL |
| Ethidium bromide | 10 µL |

7. Primer preparation (WT1 primers and GAPDH primers)

- WT1 (1) sense primer :5'- GGCATCTGAGACCAGTGAGAA-3'

352.63 µg (26 µg/OD), OD 13.40

MW : 6505.2 µg/ µmole, 54.3 nmole

- WT1 (2) anti-sense primer :5'- GAGAGTCAGACTTGAAAGCAGT-3'

268.79 µg (26 µg/OD), OD 10.40

MW : 6833.4 µg/ µmole, 39.3 nmole

- GAPDH sense primer : 5'-CGAAGTCAACGGATTTGGTCGTAT-3'

313.86 µg (28 µg/OD), OD 11.40

MW : 7408.8 µg/ µmole, 42.4 nmole

- GAPDH anti-sense primer :5'- AGCCTTCTCGGTGGTGAAGAC -3'

321.34 μg (28 $\mu\text{g}/\text{OD}$), OD 11.30

MW : 6463.2 $\mu\text{g}/\mu\text{mole}$, 49.7 nmole

Dissolve primers in 1 mL of sterile deionized distilled water, the concentration of primer solution of WT1(1), WT1(2), GAPDH sense primer and GAPDH anti-sense primer in stock 1 will be 54.3, 39.9, 42.4 μM , respectively. Kept at -20°C

Stock solution 2 should be diluted from the stock solution 1 in the final concentration of 10 μM in sterilized deionized distilled water.

8. Component of RT-PCR

For K562, HL60 and Molt4 cell line

| Components | Volume/ 20 μL | Final concentration |
|---------------------------|--------------------------|---------------------|
| 2X reaction mix | 10 μL | 1X |
| Template RNA | X μL * | 1 μg |
| WT1 (1) sense primer | 0.4 μL | 0.2 μM |
| WT1 (2) anti-sense primer | 0.4 μL | 0.2 μM |
| GAPDH Sense primer | 1.7 μL | 0.085 μM |
| GAPDH anti-sense primer | 1.7 μL | 0.085 μM |
| RT-Taq Mix | 0.4 μL | - |
| DEPC treated water | up to 20 μL | - |

For U937 cell line

| Components | Volume/ 20 μL | Final concentration |
|---------------------------|--------------------------|---------------------|
| 2X reaction mix | 10 μL | 1X |
| Template RNA | X μL * | 1 μg |
| WT1 (1) sense primer | 0.6 μL | 0.3 μM |
| WT1 (2) anti-sense primer | 0.6 μL | 0.3 μM |
| GAPDH Sense primer | 1.7 μL | 0.085 μM |
| GAPDH anti-sense primer | 1.7 μL | 0.085 μM |
| RT-Taq Mix | 0.8 μL | - |
| DEPC treated water | up to 20 μL | - |

* The volume of template RNA varies to the amount of RNA in each of cell

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