



APPENDICES

ลิขสิทธิ์มหาวิทยาลัยเชียงใหม่
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APPENDIX A

Reagents and buffer preparation

Reagents for density gradient ultracentrifugation

1. Density solution 1.006 g/mL (solution A)

- NaCl	22.8	g
- Na ₂ EDTA	0.2	g
- NaOH (1mol/L)	2	ml
- Double distilled H ₂ O to	2	L

2. Density solution 1.019 g/mL

- KBr	0.376	g
- solution A to	20	ml

3. Density solution 1.063 g/mL

- KBr	1.682	g
- solution A to	20	ml

4. Density solution 1.24 g/mL

- KBr	7.497	g
- solution A to	20	ml

Reagents for dialysis

1. 10X PBS buffer, pH 7.2

- NaCl	80	g
- KCl	2	g
- Na ₂ HPO ₄ (MW 141.96)	11.5	g
- KH ₂ PO ₄ (MW 174.18)	2	g
- Double distilled H ₂ O to	1,000	ml

Reagents for Lowry protein assay

1. Reagent A

- CTC	5	ml
- 20% Na ₂ CO ₃	5	ml
- 0.8 N NaOH	10	ml
- 5% SDS	20	ml

2. Reagent B

- Folin-Ciocalteu phenol reagent	1	ml
- Sterile water	5	ml

3. Stock solution CTC

- CuSO ₄ .7H ₂ O	0.2	g
- Tatalic acid	0.4	g
- Sterile water to	100	ml

4. 0.8 N NaOH

- NaOH	1.6	g
- Sterile water to	50	ml

5. 5% SDS

- SDS	2.5	g
- Sterile water to	50	ml

6. 20% Na₂CO₃

- Na ₂ CO ₃	10	g
- Sterile water to	50	ml

Reagents for one-dimensional electrophoresis**1. 12.5% acrylamide solution**

- 40% acrylamide solution	3.125	ml
- 1.5 M Tris-HCl pH 8.8	2.5	ml
- 10% SDS	125	μl
- Double distilled water	4.225	ml
- 10% APS	50	μl
- TEMED	3.3	μl

2. stacking solution

- 40% acrylamide solution	300	μl
- 0.5 M Tris-HCl pH 6.8	742	μl
- 10% SDS	30	μl
- Double distilled water	1.916	ml
- 10% APS	23	μl
- TEMED	1.7	μl

3. 1.5 M Tris-HCl pH 8.8

- Tris 36.3 g
- HCl to pH 8.8
- Double distilled water to 200 ml

4. 0.5 M Tris-HCl pH 6.8

- Tris 3 g
- HCl to pH 6.8
- Double distilled water to 50 ml

5. 10% SDS solution

- SDS 10 g
- Double distilled water to 100 ml

6. SDS electrophoresis buffer

- Tris 3.02 g
- Glycine 14.4 g
- SDS 1 g
- Double distilled water to 1000 ml

7. 10% ammonium persulfate solution (10% APS)

- Ammonium persulfate 0.1 g
- Double distilled water to 1 ml

8. Silver staining reagents

8.1 Fixing solution

- 50% Methanol	500	ml
- 12% Acetic acid	120	ml
- 37% Formaldehyde	500	µl
- Double distilled water to	1000	ml

8.2 Washing solution

- Ethanol	350	ml
- Double distilled water to	1000	ml

8.3 Sensitizing solution

- Sodium thiosulfate	0.2	g
- Double distilled water to	1000	ml

8.4 Staining solution

- Silver nitrate	2	g
- Double distilled water to	1000	ml

8.5 Developing solution

- Sodium carbonate	60	g
- 0.02% Sodium thiosulfate	20	ml
- 37% Formaldehyde	500	µl
- Double distilled water to	1000	ml

8.6 Stopping solution

- Sodium-EDTA	14.6	g
- Double distilled water to	1000	ml

8.7 Storing solution

- Acetic acid 1 ml
- Double distilled to 1000 ml

Reagent for two-dimension electrophoresis

1. Lysis solution (7M urea, 2M thiourea, 4% CHAPS)

- Urea 4.2 g
- Thiourea 1.5 g
- CHAPS 0.4 g
- Double distilled water to 10 ml

2. Rehydration stock solution without IPG buffer

- Urea 4.2 g
- Thiourea 1.5 g
- CHAPS 0.4 g
- Bromphenol blue 20 µl of 1% solution
- Double distilled water to 10 ml

3. SDS equilibration buffer

- Tris-HCl pH 8.8 10 ml
- Urea 72.07 g
- Glycerol 69 ml
- SDS 4 g
- Bromphenol blue 400 µl of 1% solution
- Double distilled water to 200 ml

4. Agarose sealing solution

- SDS electrophoresis buffer 10 ml
- Agarose 0.05 g
- Bromphenol blue 20 µl of 1% solution

5. Bromphenol blue stock solution

- Bromphenol blue 10 mg
- Tris-base 6 mg
- Double distilled water to 1 ml

Reagents for In-gel trypsin digestion

1. 20 mM Ammonium bicarbonate

- Ammonium bicarbonate 79.6 mg
- Sterile water to 50 ml

2. 10 mM DTT/ 10 mM ammonium bicarbonate

- DTT 7.7125 mg
- 10 mM ammonium bicarbonate 5 ml

3. 100 mM Iodoacetamide/ 10 mM ammonium bicarbonate

- Iodoacetamide 92 mg
- 10 mM ammonium bicarbonate 5 ml

4. 10 ng Trypsin in 50% acetonitrile/ 10 mM ammonium bicarbonate

- Trypsin 20 µg
- 50% acetonitrile/ 10 mM ammonium bicarbonate 2 ml

5. 30% acetonitrile

- 100% acetonitrile 1.5 ml
- Sterile water to 5 ml

6. 0.1% FA

- Formic acid 50 μ l
- Sterile water to 50 ml

7. 50% acetonitrile/ 0.1% FA

- 100% acetonitrile 5 ml
- 0.1% FA 5 ml

APPENDIX B

Condition for MS/MS ions search

MASCOT MS/MS Ions Search

Your name	keeratiporn	Email	sessionalpaper@hotmail.com
Search title	1 b		
Database(s)	Environmental_EST SwissProt NCBInr contaminants cRAP	Enzyme	Trypsin
		Allow up to	1 missed cleavages
		Quantitation	None
Taxonomy Homo sapiens (human)		
Fixed modifications	Carbamidomethyl (C)	>	Acetyl (K) Acetyl (N-term) Acetyl (Protein N-term) Amidated (C-term) Amidated (Protein C-term) Ammonia-loss (N-term C) Biotin (K) Biotin (N-term) Carbamyl (K) Carbamyl (N-term) Carboxymethyl (C)
	Display all modifications		
Variable modifications	Dioxidation (M)	>	
<		<	
Peptide tol. ±	1.2 Da	# ^{13}C 0	MS/MS tol. ± 0.6 Da
Peptide charge	1+, 2+ and 3+	Monoisotopic <input checked="" type="radio"/> Average <input type="radio"/>	
Data file	D:\ทีมงานพัฒนา\biotec\2D 5 march 2011\result...		
Data format	Mascot generic	Precursor	m/z
Instrument	ESI-TRAP	Error tolerant	<input type="checkbox"/>
Decoy	<input type="checkbox"/>	Report top	50 hits
Start Search ...		Reset Form	

Figure 28 MASCOT MS/MS ions search

CURRICULUM VITAE

Name Miss Keeratiporn Worrasettasing

Date of Birth February 21, 1987

Education background

Year	Degree	Institution
2005	Certificated of high school	Yothinburana
2009	Bachelor of Science (Medical Technology)	Chiang Mai University

Research support

2011 The NSTDA Research Chair Grant
 The National Research University Project under Thailand's
 Office of the Higher Education Commission
 PHC Franco-Thai cooperation program in higher education and
 research
 Graduate school, Chaing Mai University

Honors and Awards

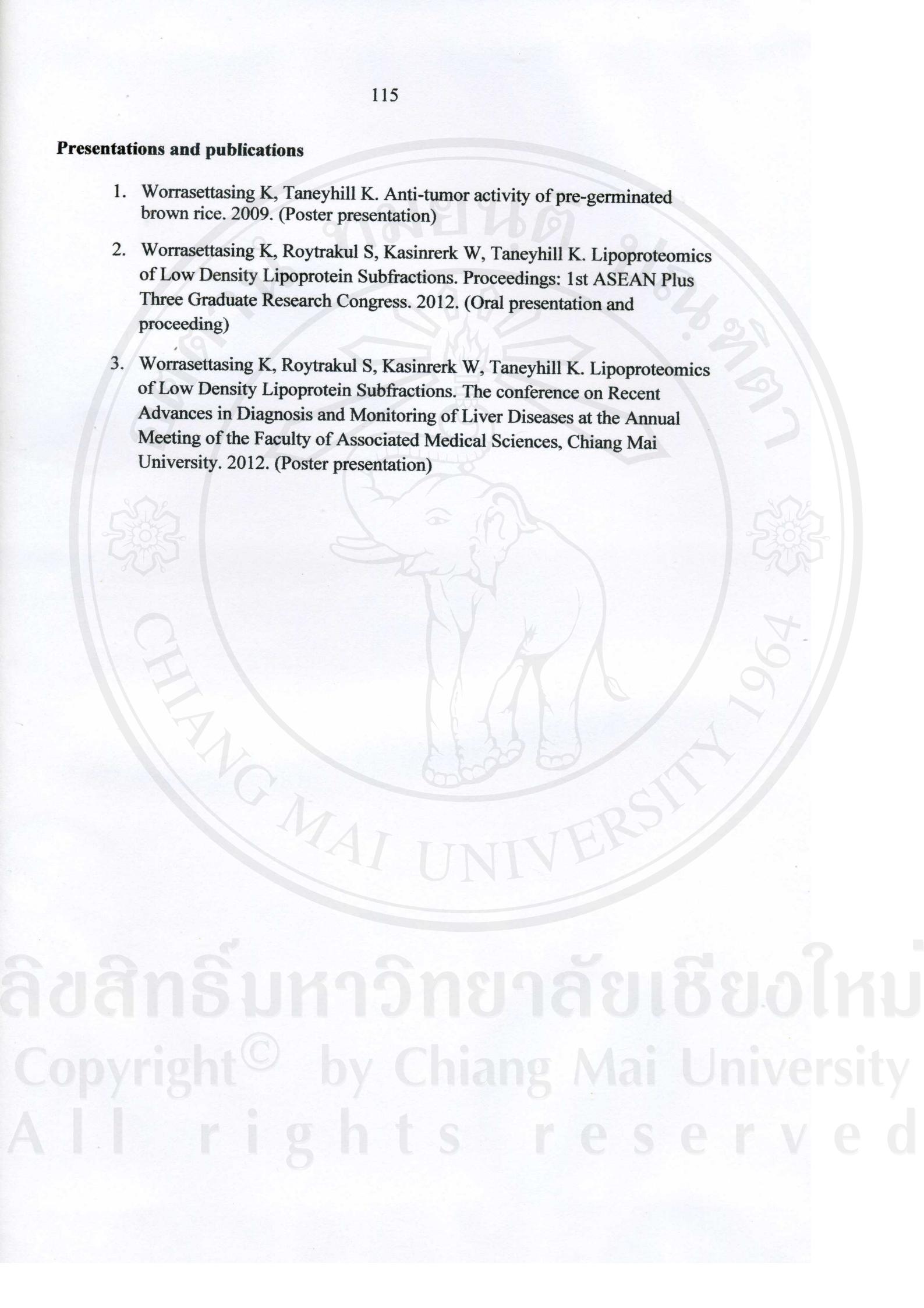
2012 Excellent Oral Presentation at The 1st ASEAN Plus Three

Graduate Research Congress (AGRC 2012) at The Empress
Hotel, Chiang Mai, Thailand

2009 Excellent Poster Presentation at Faculty of Associated Medical
Sciences, Chiang Mai University

Presentations and publications

1. Worrasettasing K, Taneyhill K. Anti-tumor activity of pre-germinated brown rice. 2009. (Poster presentation)
2. Worrasettasing K, Roytrakul S, Kasinrerk W, Taneyhill K. Lipoproteomics of Low Density Lipoprotein Subfractions. Proceedings: 1st ASEAN Plus Three Graduate Research Congress. 2012. (Oral presentation and proceeding)
3. Worrasettasing K, Roytrakul S, Kasinrerk W, Taneyhill K. Lipoproteomics of Low Density Lipoprotein Subfractions. The conference on Recent Advances in Diagnosis and Monitoring of Liver Diseases at the Annual Meeting of the Faculty of Associated Medical Sciences, Chiang Mai University. 2012. (Poster presentation)



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